



SasView Tutorials

**Basic 1D Data Fitting
in SasView Version 6**

www.sasview.org

Preamble

SasView was originally developed by the University of Tennessee as part of the Distributed Data Analysis of Neutron Scattering Experiments (DANSE) project funded by the US National Science Foundation (NSF), but is currently being developed as an Open Source project hosted on GitHub and managed by a consortium of scattering facilities. Participating facilities include (in alphabetical order): the Australian National Science & Technology Centre for Neutron Scattering, the Diamond Light Source, the European Spallation Source, the Federal Institute for Materials Research and Testing, the Institut Laue Langevin, the ISIS Pulsed Neutron & Muon Source, the National Institute of Standards & Technology Center for Neutron Research, the Oak Ridge National Laboratory Neutron Sciences Directorate, and the Technical University Delft Reactor Institute.

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If you make use of SasView

If you use SasView to do productive scientific research that leads to a publication, we ask that you acknowledge use of the program with the following text:

This work benefited from the use of the SasView application, originally developed under NSF Award DMR-0520547. SasView also contains code developed with funding from the EU Horizon 2020 programme under the SINE2020 project Grant No 654000.

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Learning Objective

This tutorial will demonstrate how to fit individual 1D ('intensity' versus Q) datasets in SasView and showcase some of the associated functionality of the program. Simultaneous or batch fitting of multiple 1D datasets, and the fitting of 2D datasets, are considered in separate tutorials.

It is assumed that the reader has some familiarity with the purpose and principles of data fitting. If not, these Wikipedia articles provide an overview:

- https://en.wikipedia.org/wiki/Curve_fitting
- https://en.wikipedia.org/wiki/Mathematical_optimization

The program interface shown in this tutorial is SasView Version 6.0.0 running on a Windows platform but, apart from a few small differences in look and functionality, this tutorial is generally applicable to any version of SasView Version 6.x on any platform. However, there are separate tutorials for SasView 5.x and for using the old program interface released with SasView 4.x.

Glossary

<i>a priori</i> information	Known facts about the system whose datasets are being modelled that can guide the selection of model or model parameters.
Chi-square (X^2 , 'Chi2')	<p>A statistical test of how well a chosen model fits the data with a given set of model parameters. In SasView this means</p> $X^2 = \sum ((I(Q)_{meas} - I(Q)_{calc})^2 / E(Q)^2)$ <p>where I is the scattering intensity and E is the error on the intensity value. Clearly, as $X^2 \rightarrow 0$, the better the model fit is.</p> <p>NB: The SasView interface actually reports a variation of chi-square called the <u>reduced chi-square</u>, sometimes referred to as the '<u>goodness-of-fit</u>'</p> $X^2_{Reduced} = \sum ((I(Q)_{meas} - I(Q)_{calc})^2 / E(Q)^2) / (N_{pts} - N_{params})$ <p>where N_{pts} is the number of data points in the dataset and N_{params} is the number of model parameters being optimised (which may be less than the total number of parameters in the model!). As $X^2_{Reduced} \rightarrow 1$, the better the model fit is.</p>
Compute/Plot [button]	Perform a direct calculation of the model with the current parameters but <u>without any optimisation</u>

Correlated parameters	<p>To obtain the best solutions the optimiser needs to explore the widest possible parameter space. That is best achieved if every parameter can be considered independent of every other parameter. However, there will be instances where this is not the case and one or more parameters may be correlated. In such cases the best approach is to fix the values of some of correlated parameters using <i>a priori</i> information and then optimise the remaining values.</p> <p>A particularly common instance of correlated parameters encountered when model-fitting SAS data is when the components of the forward scattering intensity</p> $I(0) = \phi V (\Delta \rho)^2$ <p>where Φ is the volume fraction of scatterers, V is the volume of one scatterer (and so dependent on size parameters), and $\Delta\rho$ is the contrast (difference in SLDs), are separated out as individual parameters in a model. Optimizing two or more of these parameters will cause them to be correlated.</p> <p>Correlated parameters often manifest themselves in the SasView interface as having very large uncertainties.</p>
Fit [button]	Perform a calculation of the model <u>optimising the selected parameters</u>
Model-fitting	The process of finding a good yet <u>physically-realistic</u> mathematical description ('solution') for a dataset or collection of datasets. The procedure employed to achieve this is called optimisation.
OpenCL	A low-level software framework that allows calculations to be distributed between any compatible processors (eg, GPUs as well as CPUs) on the host computer. OpenCL can speed up demanding model-fits if suitable hardware is available.
OpenMP	A software framework that permits shared-memory multi-processing (ie, parallelisation) of calculations. OpenMP can speed up demanding model-fits if suitable hardware is available.
Optimiser	The mathematical algorithm used to perform the model-fitting.
(Poly)dispersity	<p>Where one or more model parameters have a distribution of values.</p> <p>SasView allows for 2 types of (poly)dispersity:</p> <p><i>Size</i> where, for example, the radii and/or lengths of the scatterers have a distribution of values <i>This will apply in most instances</i></p>

	<p><i>Angular</i> where the scatterers are anisometric in shape (eg, cylinders) and exhibit preferred orientations with respect to Q, for example, under shear or in a magnetic field</p>
Reduced Chi2	See Chi-square
Residual	<p>The difference between the measured and calculated values of a function at a given point. In SasView this means:</p> $R(Q) = I(Q)_{meas} - I(Q)_{calc}$ <p>where I is the scattering intensity.</p> <p>NB: The SasView interface actually reports a variation of the residual called the <u>normalised residual</u></p> $R(Q)_{Normalised} = (I(Q)_{meas} - I(Q)_{calc}) / E(Q)$ <p>where E is the error on the intensity value.</p> <p>A normalised residual can be thought of as the number of standard deviations between the measured value and the calculated value. Thus, for a good fit, 68% of the values will lie within $-1 < R(Q)_{Normalised} < +1$, and 95% within ± 2.</p> <p>Residuals larger than ± 3 indicate significant problems with either the input data or the choice of model or model parameters.</p>
SLD	<p>Abbreviation for Scattering Length Density, a measure of the ability of a molecule to scatter. Strictly speaking, SLD is a SANS quantity, so if fitting SAXS data use electron density values in their place.</p> <p>SLD values (neutron and X-ray) can be calculated with the SLD Calculator Tool in SasView.</p>
Smearing	<p>Sometimes the instrumental geometry used to acquire the experimental data has an impact on the clarity of features in the scattering curve. For example, peaks or fringes might be slightly broadened. This is known as Q-resolution smearing.</p> <p>To compensate for this effect SasView can add a resolution contribution into a model calculation (which by definition will be exact) to make it more representative of what has been measured experimentally.</p> <p>SasView provides 4 smearing options:</p>

	<p><i>None</i> no smearing correction is applied</p> <p><i>Use dQ Data</i> the measured Q-resolution for each data point is used to apply a smearing correction <i>This is the default if dQ data is present</i></p> <p><i>Custom Pinhole</i> SasView will apply a smearing correction suitable for data measured on 'pinhole geometry' instruments (eg, most SAXS/SANS instruments)</p> <p><i>Custom Slit</i> SasView will apply a smearing correction suitable for data measured on 'slit geometry' instruments (eg, most USAXS/USANS instruments)</p>
Theory	The name SasView gives to a model calculation.
Uncertainties	<p>Every experimental measurement, including the measurement of $I(Q)$, is subject to some degree of error (which will, ideally, be included in the dataset). Similarly, the parameters returned by optimisation will have some associated range of uncertainty.</p> <p>Parameters with uncertainties that are more than 95% of the parameter value should be viewed with deep suspicion.</p> <p>NB: Unhelpfully, but for reasons of space, the SasView interface actually labels parameter uncertainties as errors.</p>
Weighting	<p>An optimiser can be instructed to pay less or more attention to data points in a dataset by changing the weighting of those data points.</p> <p>SasView provides 4 weighting options:</p> <p><i>No Weighting</i> all data points will be weighted equally</p> <p><i>Use dI Data</i> the data points will be inversely weighted according to their measured intensity errors (ie, less prominence will be given to data points with large errors) <i>This is the default if dI data is present</i></p> <p><i>Use sqrt (I Data) </i> the data points will be inversely weighted according to the square root of their intensity values</p> <p><i>Use (I Data) </i> the data points will be inversely weighted according to their intensity values</p>

Running SasView

Windows

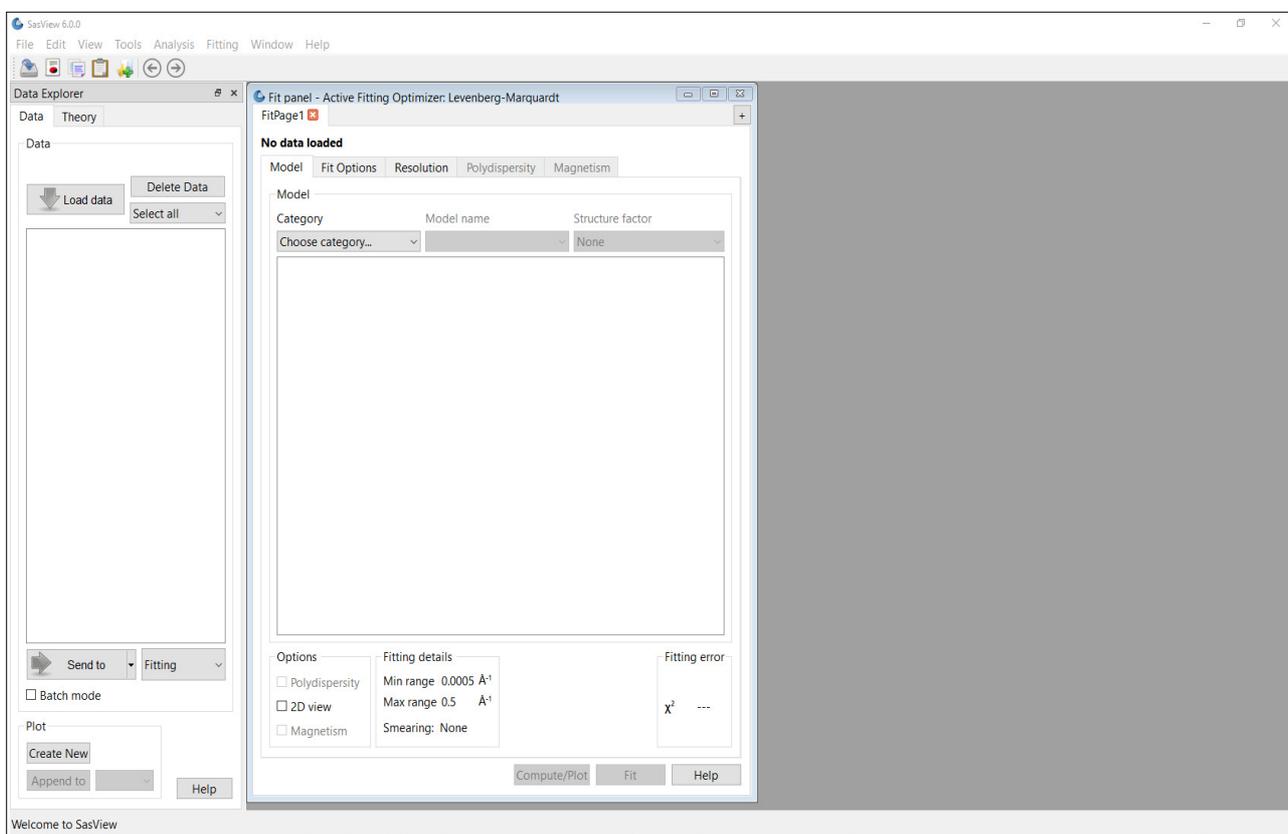
Either select SasView from 'Start' > 'All Programs' or, if you asked the installer to create one, double-click on the SasView desktop icon.



SasView

Mac OS

Go in to your 'Applications' folder and select SasView.

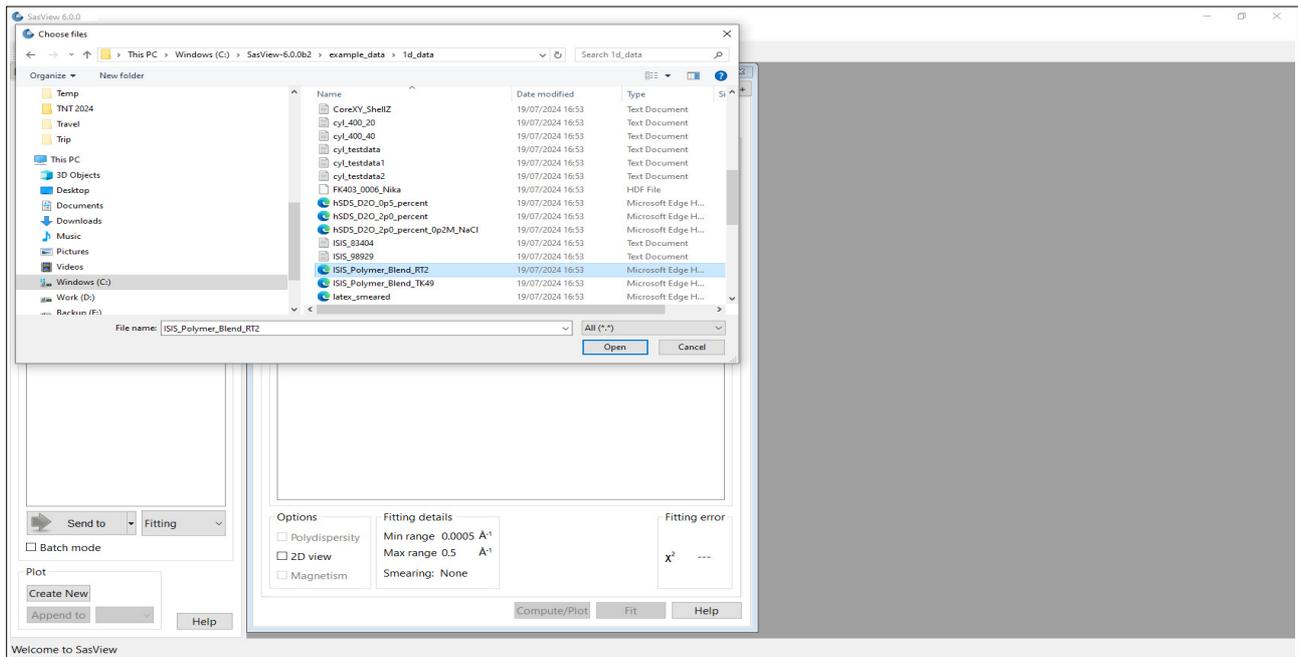


Example 1

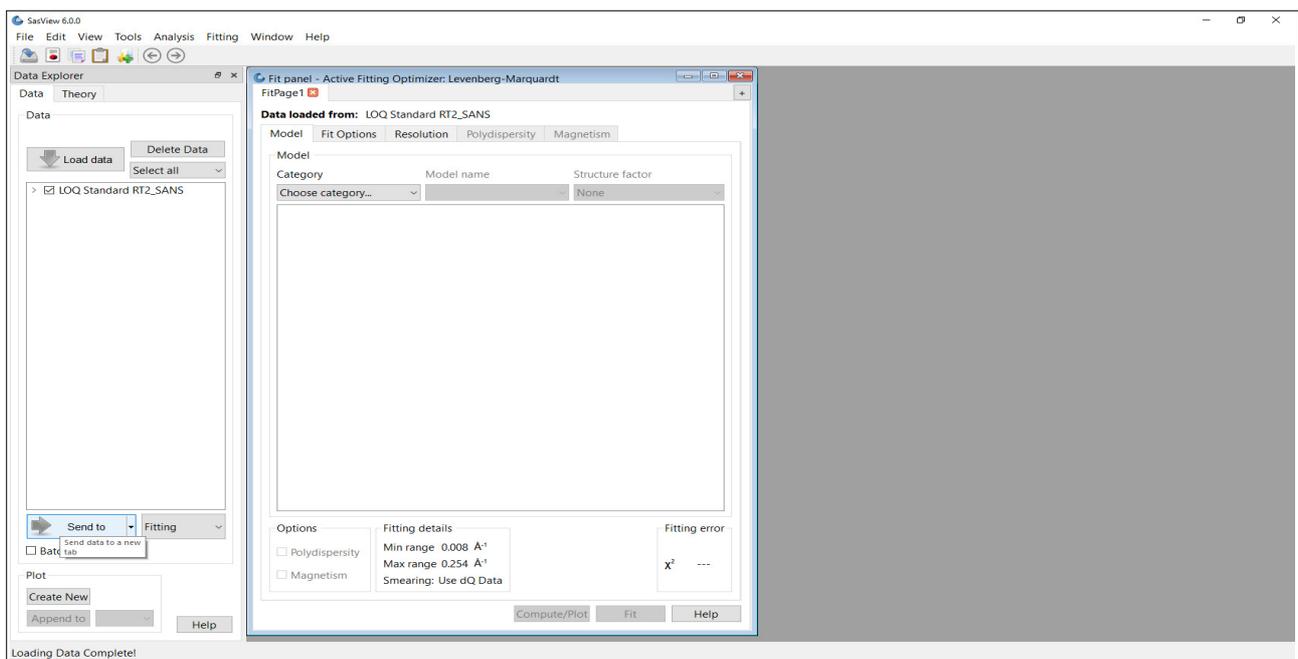
This demonstrates a simple model fit, including the impact of correlated parameters and resolution smearing.

In the Data Explorer panel, click the **Load Data** button, and navigate to the `example_data\1d_data` folder in the SasView installation directory.

Select the `ISIS_Polymer_Blend_RT2.xml` dataset and click the **Open** button.

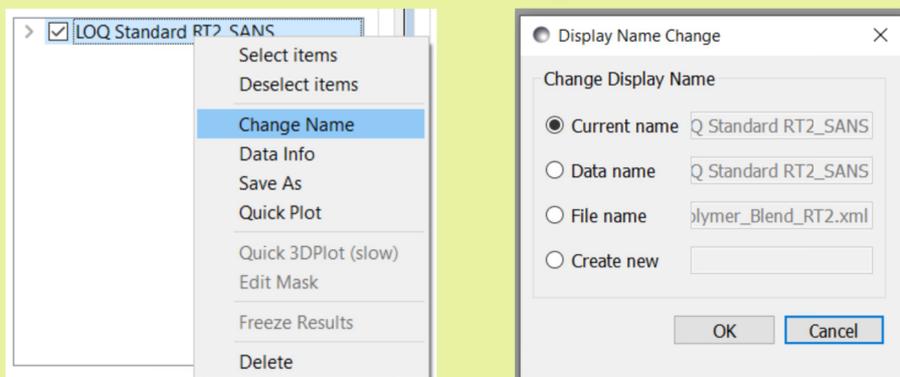


At the bottom of the Data Explorer panel, click the **Send to** button. Fitting is the default.



Tip: Notice that the name of the *data* that appears in the Data Explorer is different to the name of the *file* that was just loaded. What is being displayed is the title of the data read from within the file. Prior to SasView 5.0.4 the file name would have been displayed. Obviously, this feature depends on the file containing suitable metadata in the first place! The CanSAS1D (.xml) and NXcanSAS (.h5) standard formats are suitably compatible.

From SasView 5.0.4 it is possible to assign custom names to the data by right-clicking on a data set in the Data Explorer and selecting **Change Name**.



This brings up a dialog box with several options as shown above.

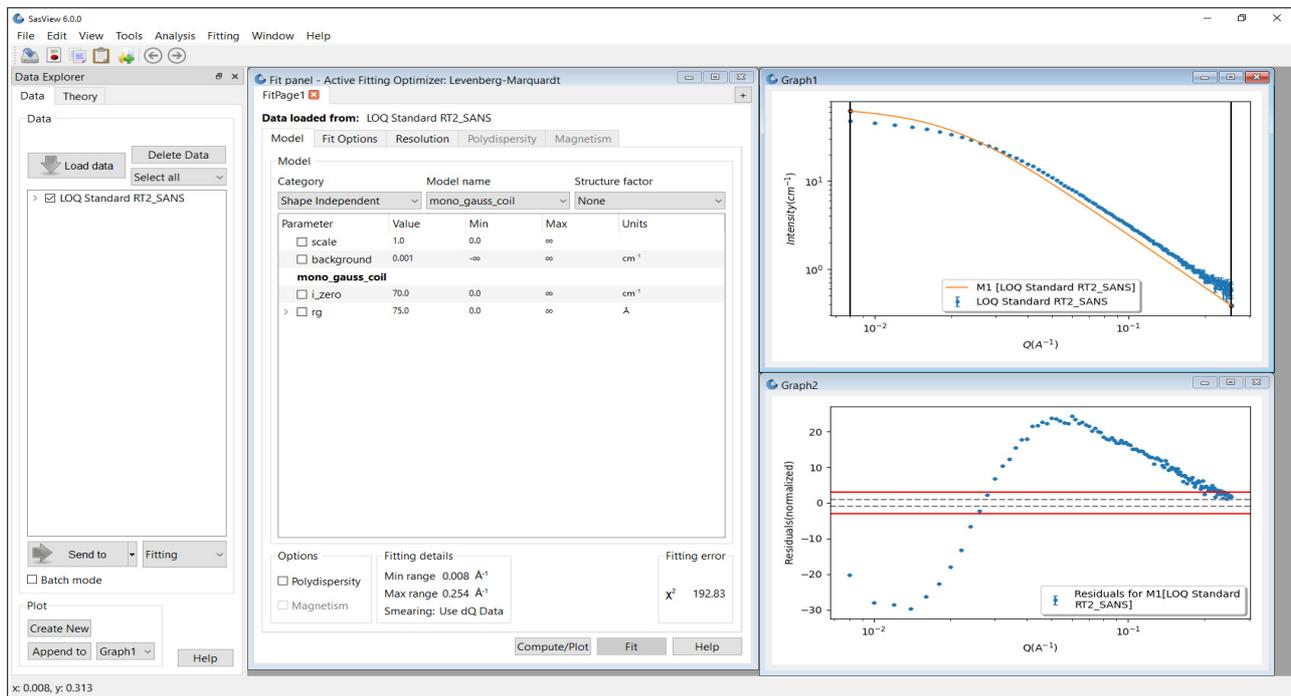
We now have to choose a model to fit to the data. To assist us in this task we have the *a priori* knowledge that this sample is a (solid) polymer blend (as it happens, of h8-polystyrene and d8-polystyrene). In the absence of solvent molecules the conformational characteristics of the polymer chains in the sample are described by a 3D random walk mediated by excluded volume constraints. Polymer physics tells us that this results in a Gaussian distribution of chain density about the centre-of-mass. The scattering from such a structure was formulated by Debye (1947) and is implemented in SasView as the **mono_gauss_coil** model.

Tip: The names of several models changed between SasView 3.x and SasView 4.x. For example, the **mono_gauss_coil** model was previously called the **Debye** model.

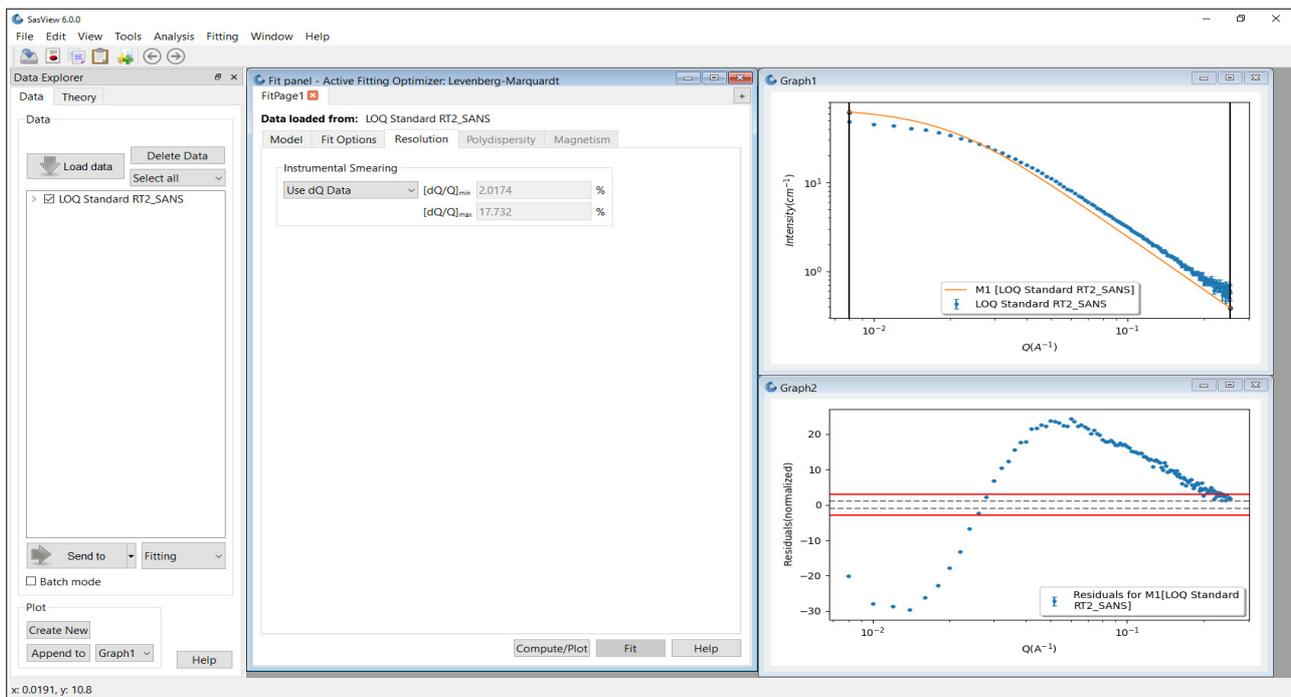
In the FitPage, select the model Category called **shape-independent** and then from the drop-down box below select the **mono_gauss_coil** model. Click the **Compute/Plot** button.

Tip: Upto and including SasView 5.0.4 the Compute/Plot button was labelled Show Plot.

The model loads with some default parameters, the calculated (not optimised at this stage) scattering from which is shown as the orange line on the upper graph. The lower graph shows the normalised residuals for the current model parameters with the boundaries for 1 (grey dashed lines) and 3 (red lines) standard deviations.



In the Resolution tab of the FitPage notice that SasView has chosen to apply resolution smearing using the measured dQ data, and in the Fit Options tab notice it will also weight the datapoints with the measured dI data. It has done this because those data are present in the dataset.

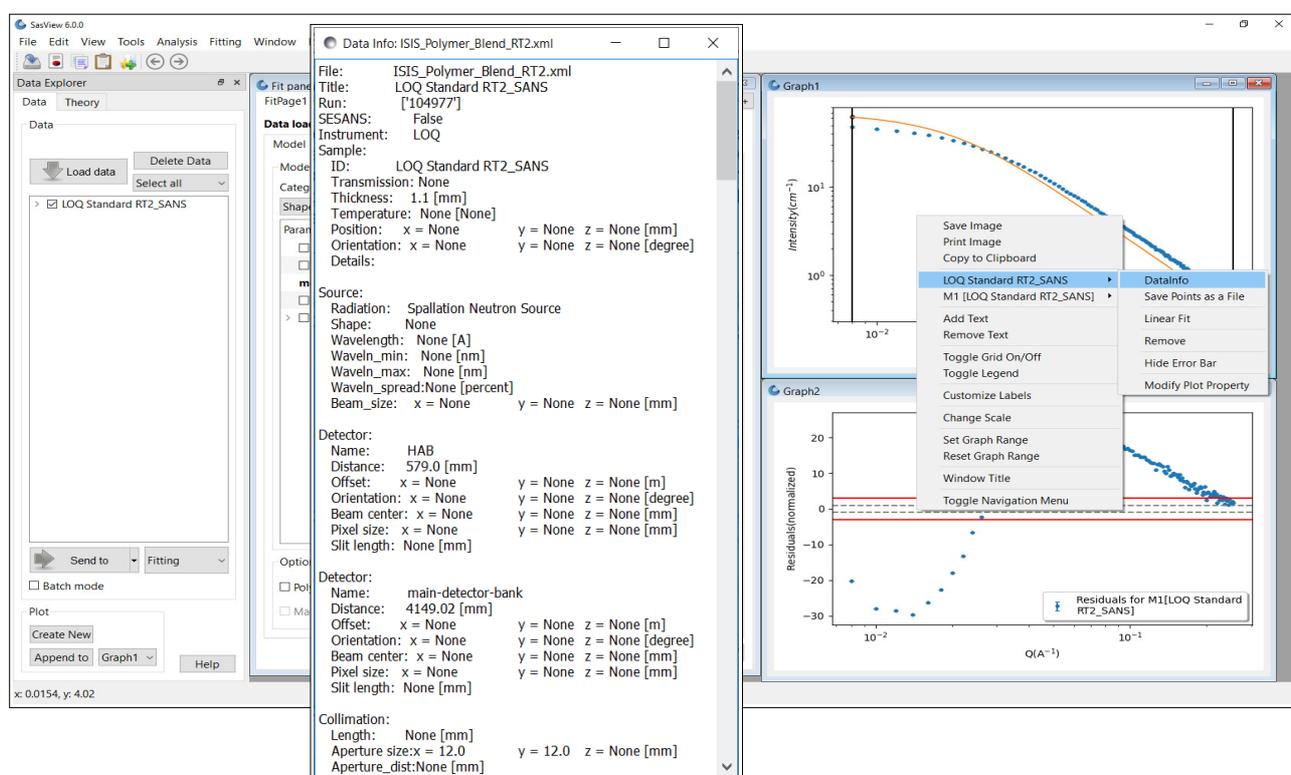


Tip: If present in the dataset, always use dI and/or dQ data to achieve a more robust solution.

By default, SasView displays all data on Log_{10} Q-axes, and scattering data on Log_{10} I-axes. To change this, right-click on a graph and select **Change Scale**. For this example, linear I-axes are better.

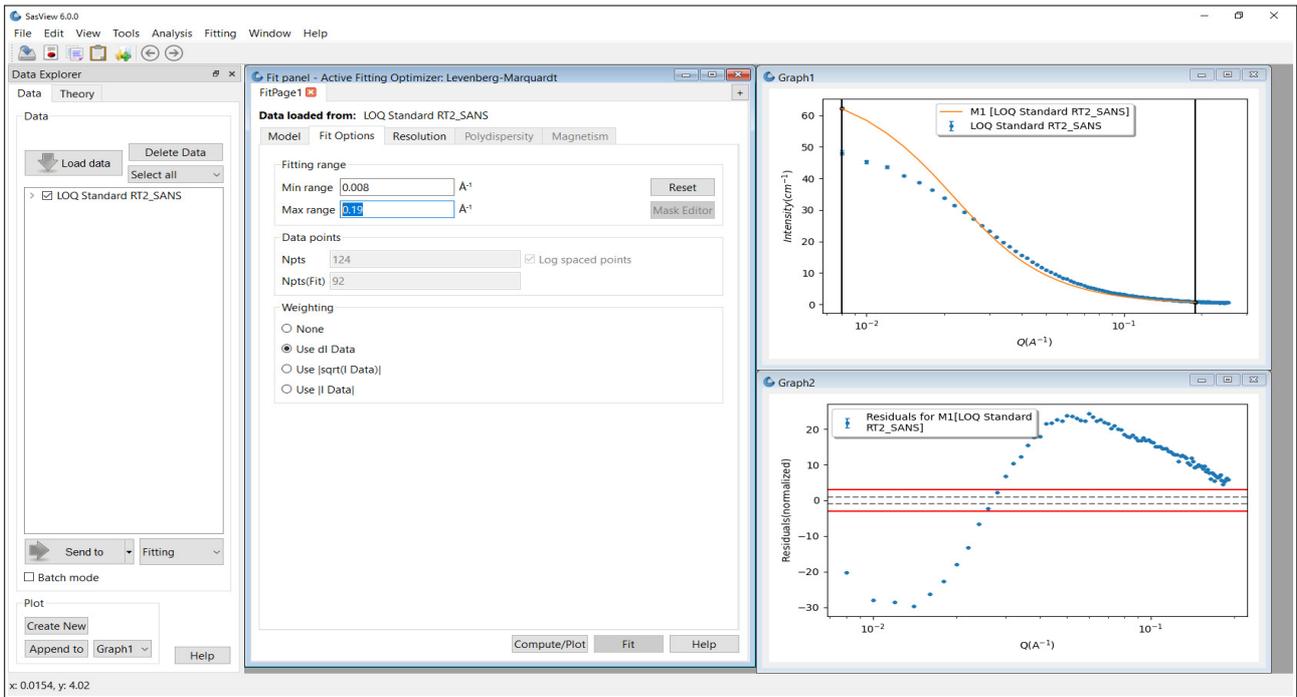
If you want to examine the contents of the dataset, right-click on the upper graph, select **ISIS_Polymer_Blend_RT2.xml** and then **DataInfo**. A separate Data Info window will appear.

The amount of information presented by DataInfo will depend on the file format of the dataset (CanSAS-standard xml files contain the most information) and the compliance of the data reduction program that wrote them.



Looking at the upper graph, it is clear that the statistical quality of the measured data degrades at higher-Q values. We can exclude these points from the fitting using the **Q range** boxes in the Fit Options tab of the FitPage.

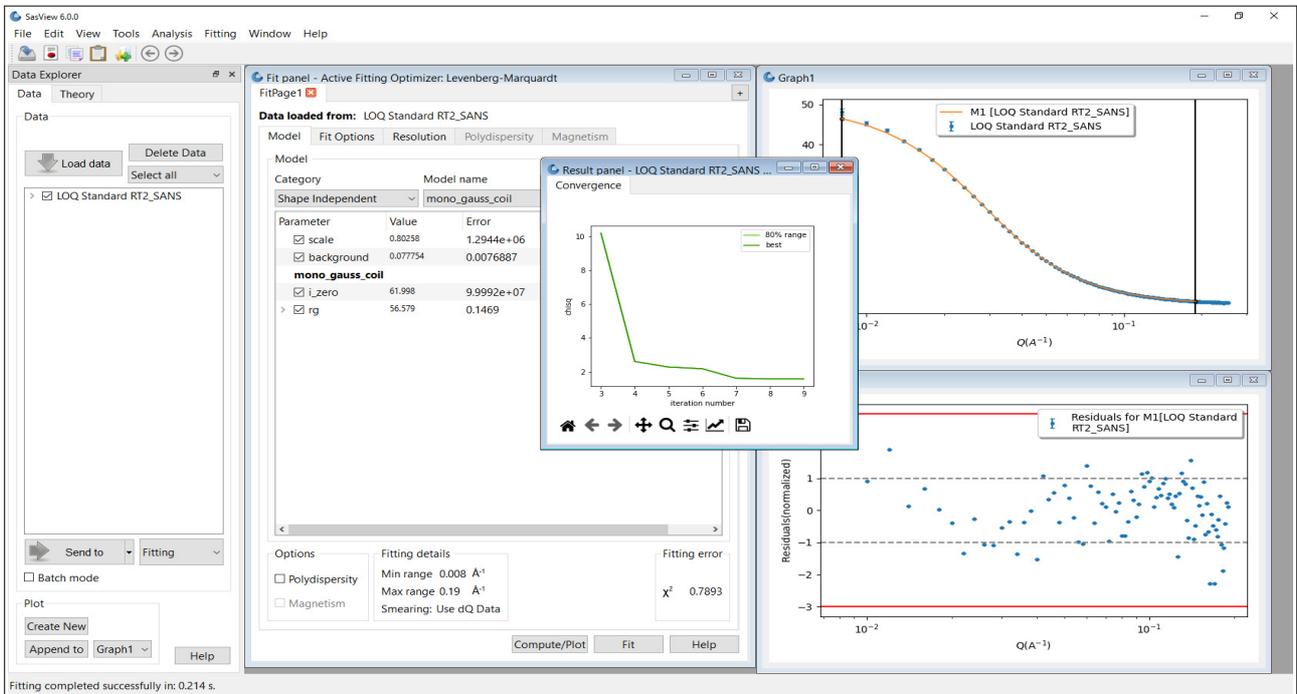
Change the Q-limits by typing values in the boxes. For this example, set the maximum-Q value to 0.19 Å⁻¹. The outer limit bar (the vertical black line on the right of the plot) moves inward to the new Q limit.



Tip: It is also possible to grab and drag the limit bars with a mouse.

Each of the 4 parameters in this model has a checkbox next to it. When the box is checked and a fit is run, the optimiser will adjust the value of the parameter.

Check all 4 boxes and then click the **Fit** button.



Several things happen very quickly:

- The progression of the fit is displayed in the Status Bar at the bottom of the SasView window
- Parameters and their uncertainties update in the FitPage
- The Reduced Chi2 (chi-squared) value updates in the FitPage
- The orange line on the upper graph updates its position and shape
- The residuals plot in the lower graph updates
- Optimiser output appears in the Log Explorer
- A Results Panel appears
- Finally, *Fitting completed successfully* appears in the Status Bar

Tip: To open the Log Explorer, select **View > Show Log Explorer**.

What do you think of the solution? It looks good, right? The model calculation ('theory' curve) is running through the measured datapoints, the normalised residuals are all within +/-2 standard deviations, and the Reduced Chi2 is close to 1 (more on this shortly!).

But, look closely at the model parameters and their uncertainties. Two of the parameters, *scale* and *i_zero*, have uncertainties (the *Error*) that make no sense. Why?

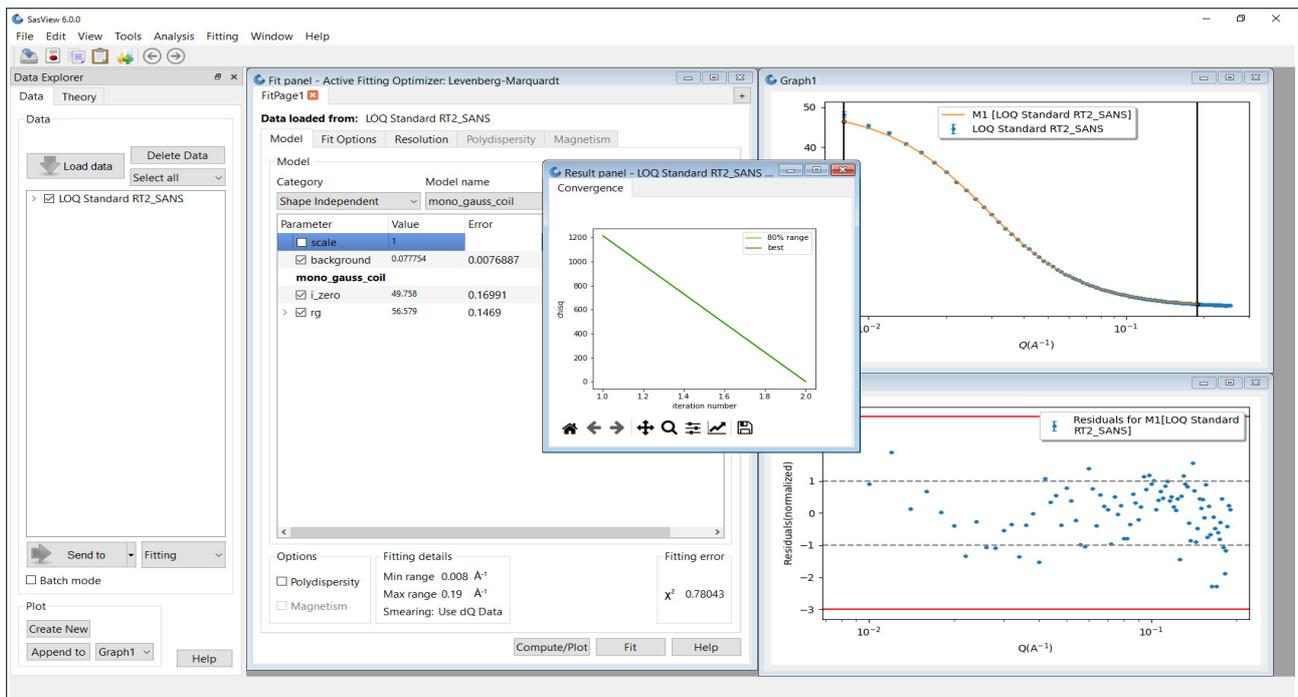
To understand this it is instructive to examine the help documentation for the model. Click the **Help** button on the FitPage (not in the Menu Bar) alongside the model name. The documentation tells us that the function that SasView is fitting is

$$I(Q) = scale \cdot I_0 \cdot P(Q) + background$$

So, in this model *scale* and *i_zero* are correlated parameters; changing one of them affects the other and the optimiser is unable to home in on a robust solution. To fit the data with this model we only need one of these parameters.

Tip: SasView adds an overall scale factor (*scale*) and a Q-independent *background* to all its models for consistency, even though some models might not need one or other of these parameters. If in doubt, consult the model help documentation.

Uncheck the *scale* parameter and set its value to 1.0. Then run the fit again (click the **Fit** button) to get a final solution.



If you want to record the solution you have a number of options:

- If you want the parameter values and their uncertainties, go to the Menu Bar and select **Edit** and either **Copy Params** (for output suitable for a text editor) or **Copy Params as...** (for output suitable for Excel or LaTeX)
 - If you want a copy of the graphs, right-click on a graph and select either **Save Image** (as .ps/eps, .jpg, .pdf, .png, .raw, .svg, or .tif), **Print Image**, or **Copy to Clipboard**
 - If you want a file of the model function (the orange line) and/or residuals, right-click on the appropriate graph and select either **M1 [LOQ Standard RT2_SANS.xml]** or **Residuals for M1 [LOQ Standard RT2_SANS.xml]** and then **Save Points as a File** (as plain ASCII text: .txt, or in CanSAS1D format: .xml)
 - If you want a two-page pdf report containing all pertinent information including copies of the graphs, go to the Menu Bar and select **Edit** and **Report Results**
- and finally
- A SasView session can be saved and reloaded as an 'Analysis' (an individual model fit), or as a 'Project' (everything done since starting your SasView session) by going to the Menu Bar and selecting **File** and either **Save Analysis** or **Save Project**.

We shall now examine some additional aspects of model-fitting this dataset.

Return to the FitPage and change the Instrumental Smearing to **None**. Then run the **Fit** again. Notice anything? The parameter values and the Reduced Chi2 value have all subtly changed. This is to be expected; Q-resolution affects the measured data, so it must have a bearing on the model fit.

Now change the model to the one called **poly_gauss_coil**. This model is an extension of Debye's original scattering law to include a Schulz-Zimm molecular weight distribution. This model has one extra parameter: the polydispersity of the polymer (M_w/M_n). Set the *polydispersity* to 1.03 (but leave it unchecked; this is more *a priori* information) and run the **Fit** again.

The parameter values change again, but there is a marked improvement (decrease) in the Reduced Chi2 value.

Lastly, reset Instrumental Smearing to **Use dQ Data** and run the **Fit** one last time.

Here is a summary of the fit results:

Param: Model	scale	bckgrd	bckgrd _err	i_zero	i_zero _err	rg	rg _err	poly disp	reduced chi2
mono use dQ	1	0.0777	0.0076	49.75	0.169	56.57	0.146	n/a	0.780
mono none	1	0.0758	0.0076	49.65	0.169	56.44	0.146	n/a	0.771
poly none	1	0.0676	0.0077	50.07	0.172	57.36	0.150	1.03	0.659
poly use dQ	1	0.0695	0.0077	50.17	0.172	57.50	0.150	1.03	0.669

So for this dataset (sample) allowing for even a tiny amount of molecular weight polydispersity – thereby maximising our use of *a priori* information - gives rise to a significant improvement in the quality of the fit! And though it does appear that including the Q-resolution smearing slightly degrades the quality of the fit in this example, it should be remembered that instrumental smearing is always present (and so should be handled) and this dataset was devoid of any fine structure (peaks, fringes).

Aside: You may have noticed that all of the Reduced Chi2 values in the summary table above are slightly *less* than 1. Whilst values >1 are indicative of a poor fit, **Reduced Chi2 values <1 are usually an indication that the data is being 'over-fit'**; for example, because the uncertainties on the data have been over-estimated (and the fact that most of the Residuals in the earlier screen shot lie within the ± 1 band is also suspicious). It is not unusual for uncertainties to be incorrect, but it is more common for them to be under-estimated, not over-estimated.

Example 2

This demonstrates a fit requiring a structure factor and introduces the use of multiple FitPages.

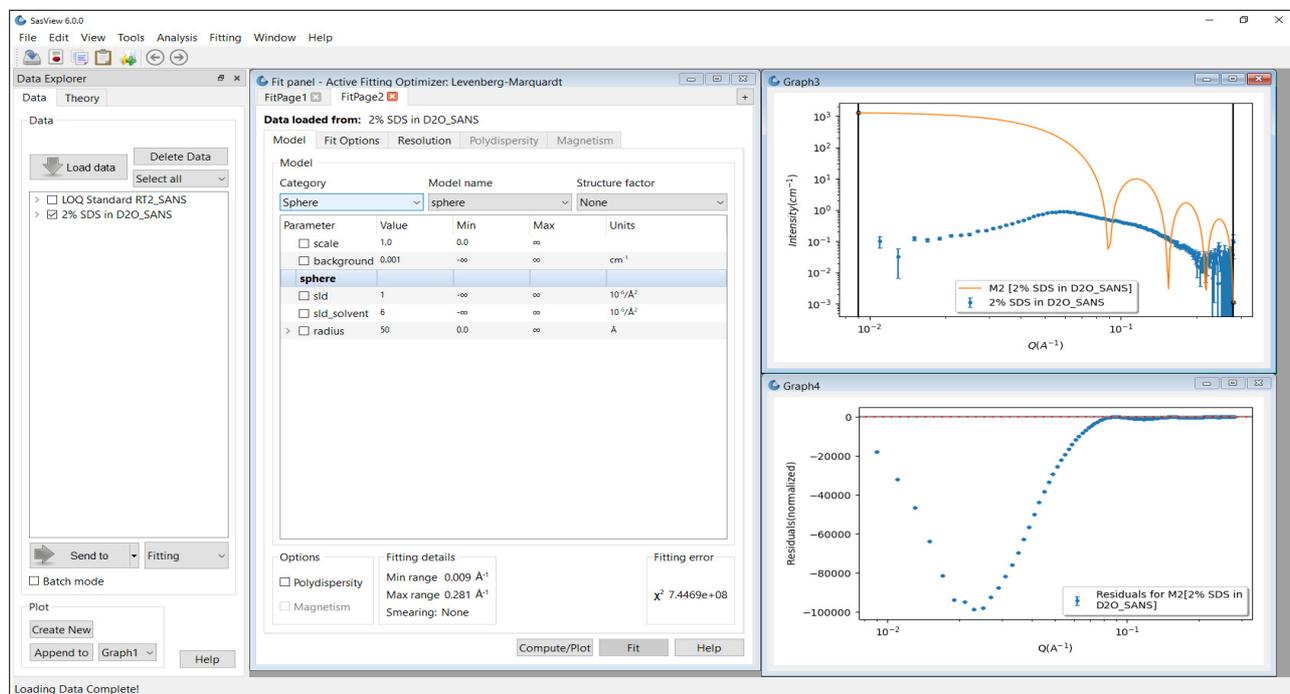
From the `\example_data\1d_data` folder in the SasView installation directory, load the `hSDS_D2O_2p0_percent.xml` dataset and send the data for fitting.

Tip: You can either: i) restart SasView, ii) click the **Delete Data** button in the Data Explorer before loading the new dataset to remove all traces of Example 1, or simply *uncheck* the LOQ Standard RT2 dataset in the Data Explorer and have SasView create a new FitPage when you click the **Send to** button, as illustrated below.

This dataset is the SANS from a 2 %^{w/w} solution of the widely-studied surfactant h25-sodium dodecyl sulphate (SDS) in heavy water (D₂O). So let us consider the available *a priori* information.

The critical micelle concentration for SDS is about 0.2 %^{w/w}, so the scattering must represent a concentrated micellar solution (though not too concentrated else the scattering would show diffraction peaks from a liquid crystalline lattice). SDS is also an anionic surfactant, so the micelles will be charged. Interactions between the charged micelles can be expected to manifest themselves in the scattering as a structure factor. And in the absence of electrolyte SDS micelles are reported to be approximately spherical.

To start with, select the **sphere** model and compute it.



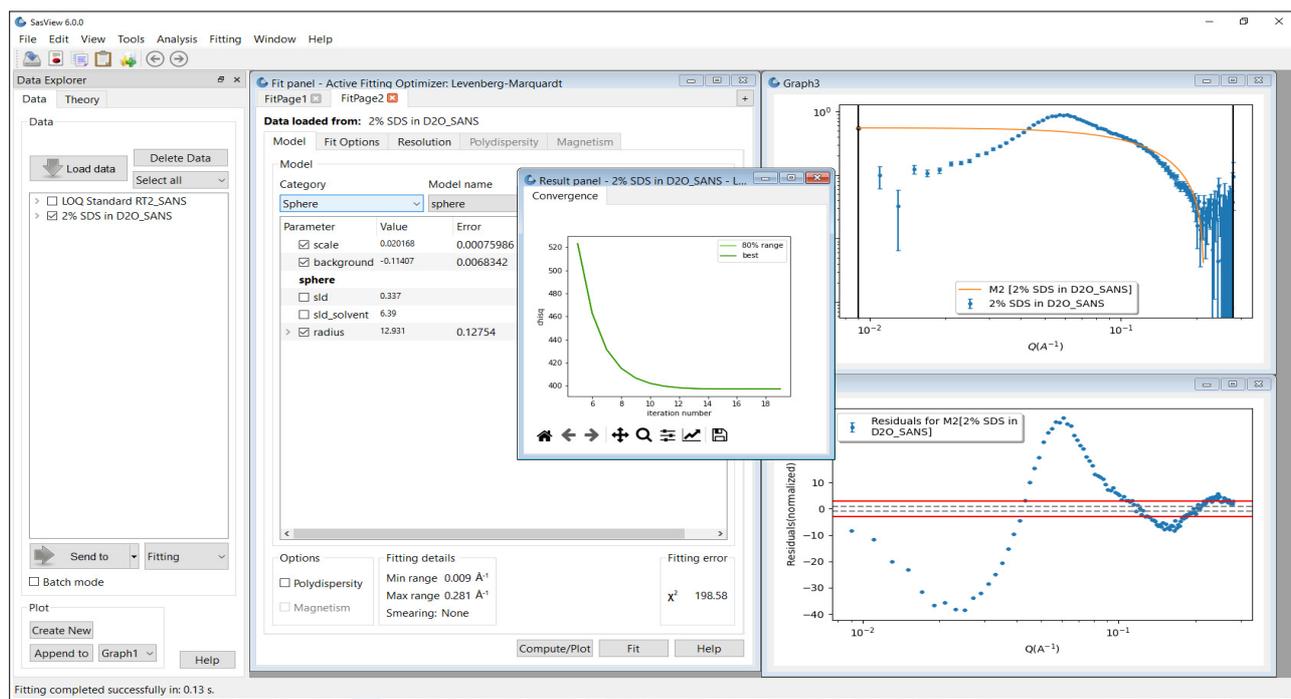
This model has 5 parameters. Two of them, *sld* (for the SDS) and *sld_solvent* (for the D₂O), can be calculated explicitly.

Go to the Menu Bar, select **Tools**, and then **SLD Calculator**. Enter the following empirical formulae and densities in the relevant boxes (leave the neutron wavelength as 6 Å):

- SDS: Na1C12H25S1O4 ; density 1.01 g/cm³
- D₂O: D2O1 ; density 1.11 g/cm³

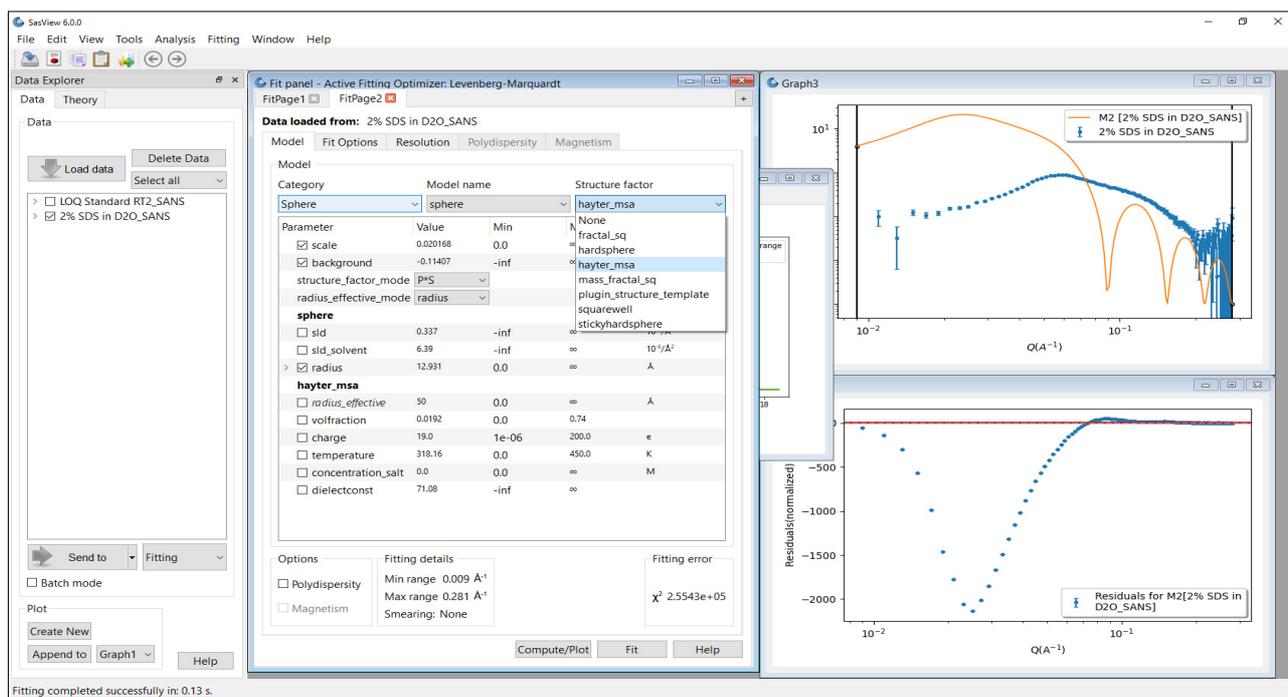
Hopefully you got values of $sld = 0.337 \times 10^{-6} \text{ \AA}^{-2}$ and $sld_{\text{solvent}} = 6.39 \times 10^{-6} \text{ \AA}^{-2}$?

Enter these values in the FitPage (they are not auto-populated from the SLD Calculator). Then check the boxes alongside *scale*, *background* and *radius* and click **Fit**.



The orange theory curve now represents the scattering from non-interacting, monodisperse, spherical micelles ~13 Å in radius. And the *scale* parameter in this case represents the volume fraction of micelles (encouragingly, about 2 %!). But to fit the broad peak – from the interacting micelles – we need to incorporate a structure factor – an $S(Q)$ – into the model.

Click on the *Structure factor* drop-down box just to the right of the $P(Q)$ *Model name* drop-down and select the **hayter_msa** $S(Q)$.



This adds 5 new parameters, but fortunately we know the values of several of them:

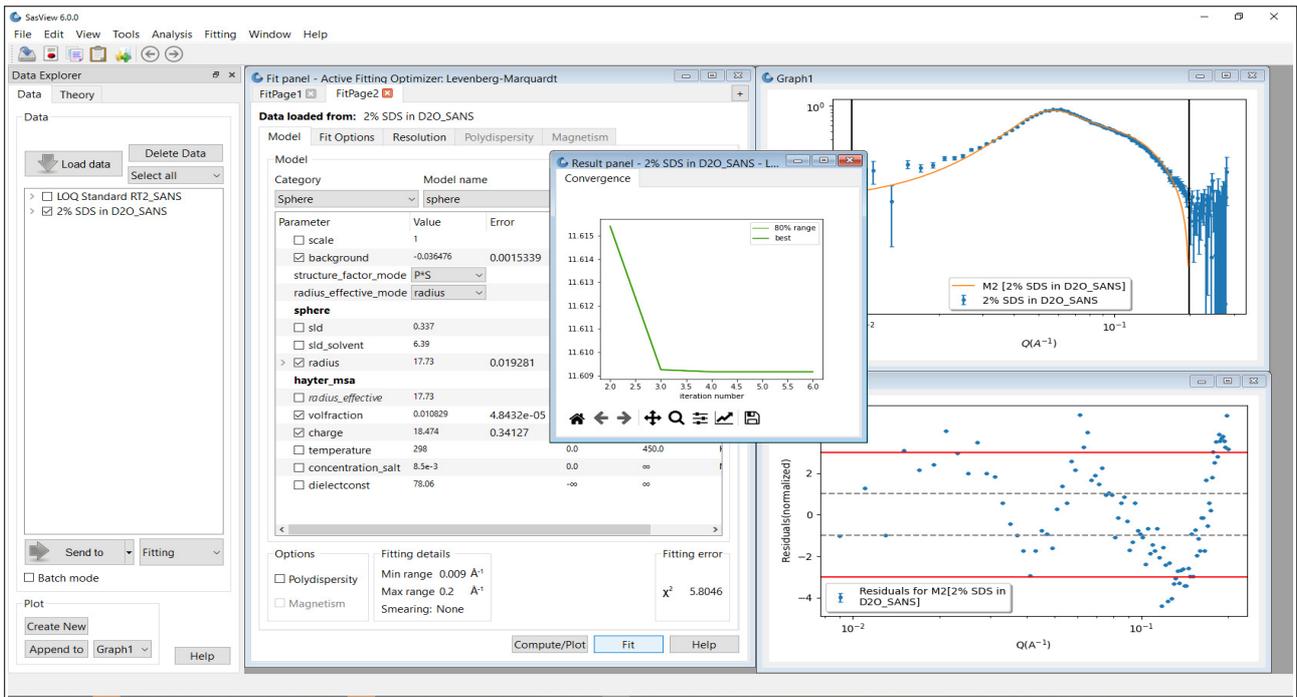
- *volfraction*: 2 %^w/_w is approximately (2/1.01) %^v/_v = 0.0198
- *charge*:
- *temperature*: 298 K
- *concentration_salt*: 8.5e-3 M
- *dielectconst*: 78.06 (Vidulich *et al*, *J Phys Chem*, 1967)

Aside: Although no additional electrolyte has been added to the system there will be dissociation of the surfactant and, therefore, dissolved ions will be present, meaning there will be a small effective salt concentration. If the micelles are in equilibrium with dissolved surfactant that is not micellised, at the critical micelle concentration the minimum ionic concentration in this particular system will likely be around 8 – 9 mM.

However, the *volfraction* parameter now duplicates the *scale* parameter. So set *scale*=1.0 and uncheck it, and check the *volfraction* parameter instead. Also check the *charge* parameter.

And also exclude some of the noisy data at high-Q by bringing the maximum fitting limit in to Q~0.2 Å⁻¹.

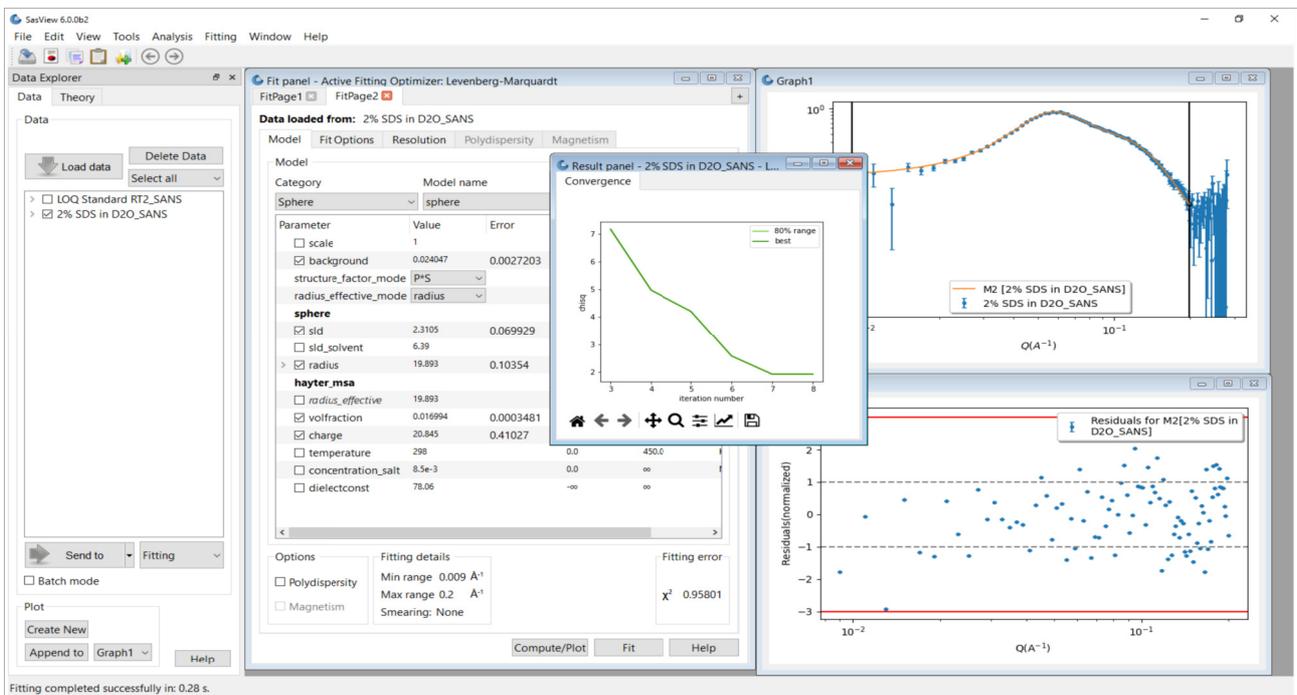
Run the fit again.



Incorporating the Hayter-Penfold MSA $S(Q)$ has demonstrably improved the fit, but there remains room for improvement: the *Reduced Chi2* is still 5.8 and the residuals range between ± 4 . Also note that the *volfraction* parameter has dropped to $\sim 1\%$.

So how might the fit be improved further? One possibility is that water molecules are mixed in with the surfactant sulphate head groups, thereby increasing the effective SLD of the surfactant molecules (and reducing the scattering contrast; ie, $slid - slid_{solvent}$).

To test this, check the *slid* parameter and run the fit again.



Indeed this does improve the fit further. The *Reduced Chi2* has dropped to ~ 1 , the residuals are within ± 2 , and the *volfraction* parameter has increased to almost 1.7 %, which is very close to the 1.78 % expected from the solution concentration (2 %) *minus* the cmc (0.2 %) = 1.8 % / 1.01.

This fit is, to all intents and purposes, as good a fit as one might reasonably expect to achieve. However, there is published evidence that suggests SDS micelles are actually slightly ellipsoidal (see, for example, DOI [10.1016/j.colsurfa.2024.134394](https://doi.org/10.1016/j.colsurfa.2024.134394) and the references therein). Can SasView confirm or refute this?

Use the **Send to** button to create another FitPage and then select the **ellipsoid** model. Do not select an S(Q) for the time being.

Set *sld* and *sld_solvent* to their calculated values.

We know from the fit to the sphere model that the micelles have a radius ~ 20 Å so set *radius_polar*=20 and *radius_equatorial*=40 (ie, to give the ellipsoids an initial axial ratio of 1:2; a sphere would be 1:1 of course). Limit the maximum Q of the fit again. Check the *scale* and *background* parameters and run the fit.

Now select the **hayter_msa** S(Q) as before and set the known values you used previously. And just like before, set *scale*=1.0 and uncheck it. But this time set the *volfraction* and *charge* parameters to the values determined from the fit to the **sphere** model; 0.017 and 20, respectively.

Also set the *radius_effective_mode* to 'equivalent_volume_sphere'.

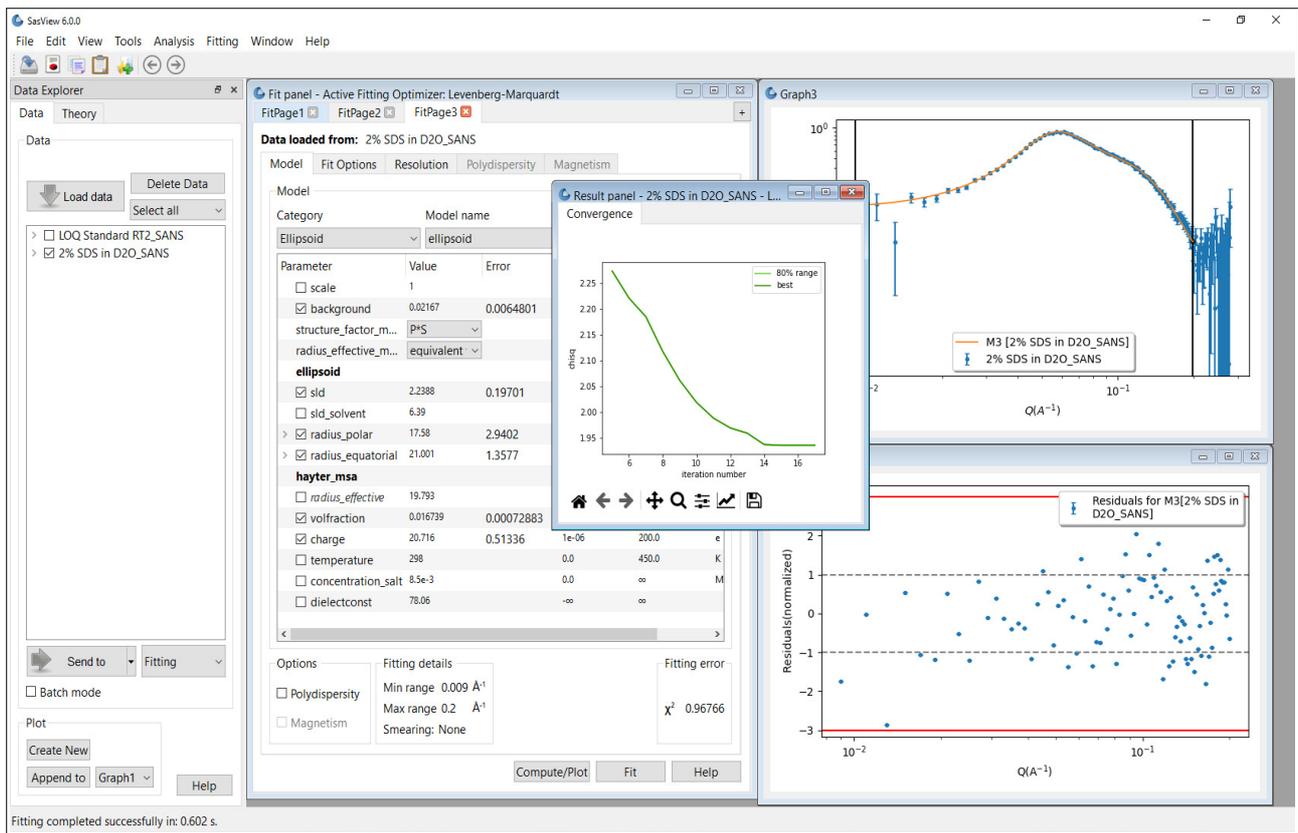
Check *radius_polar* and run the fit.

Uncheck *radius_polar*, check *radius_equatorial* and run the fit.

Now also check *radius_polar* and run the fit.

Now check *volfraction* and *charge* and run the fit.

Finally, check *sld* and run the fit.



A fit at least as good as the one to the sphere model is obtained, suggesting that the micelles are ellipsoids with an axial ratio of 1:1.2. So the SasView fitting is certainly consistent with the literature reports. But unambiguously distinguishing between the two cases would require more precise measurements and higher quality data. Just because a model fits the experimental data is not always proof that it is the correct model!

One could also explore the extent to which *size polydispersity* (see Example 3) impacts both models, or whether more nuanced models, for example, the **core_shell_sphere** or the **core_shell_ellipsoid**, allowing for a difference in SLD between the head and tail regions of the micelles, are better descriptions of the data.

Example 3

This demonstrates a fit requiring a structure factor and polydispersity.

From the `\example_data\1d_data` folder in the SasView installation directory, load the **Ludox_silica.xml** dataset and send the data for fitting.

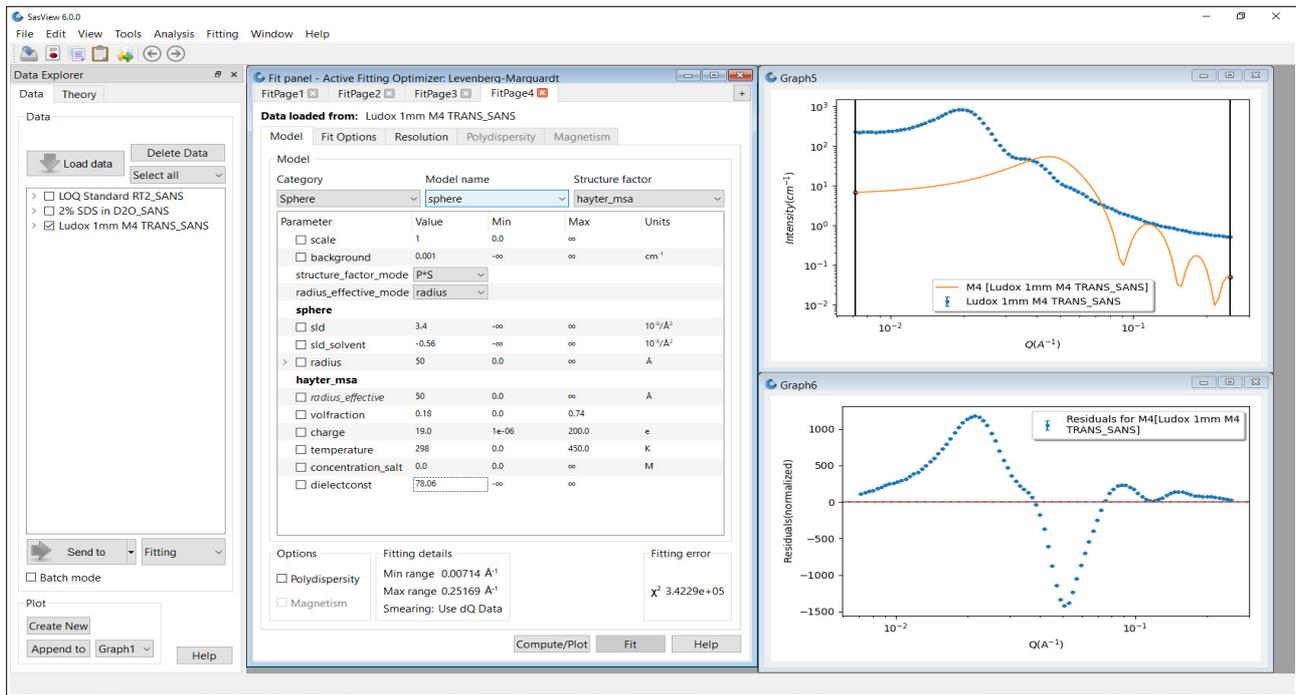
This dataset is the SANS from a concentrated dispersion of Ludox® silica nanoparticles in light water (H₂O), so there will once again be an S(Q) contribution from charged spheres.

Select the **sphere** P(Q) model and the **hayter_msa** S(Q) model.

Once again, we have a lot of *a priori* information:

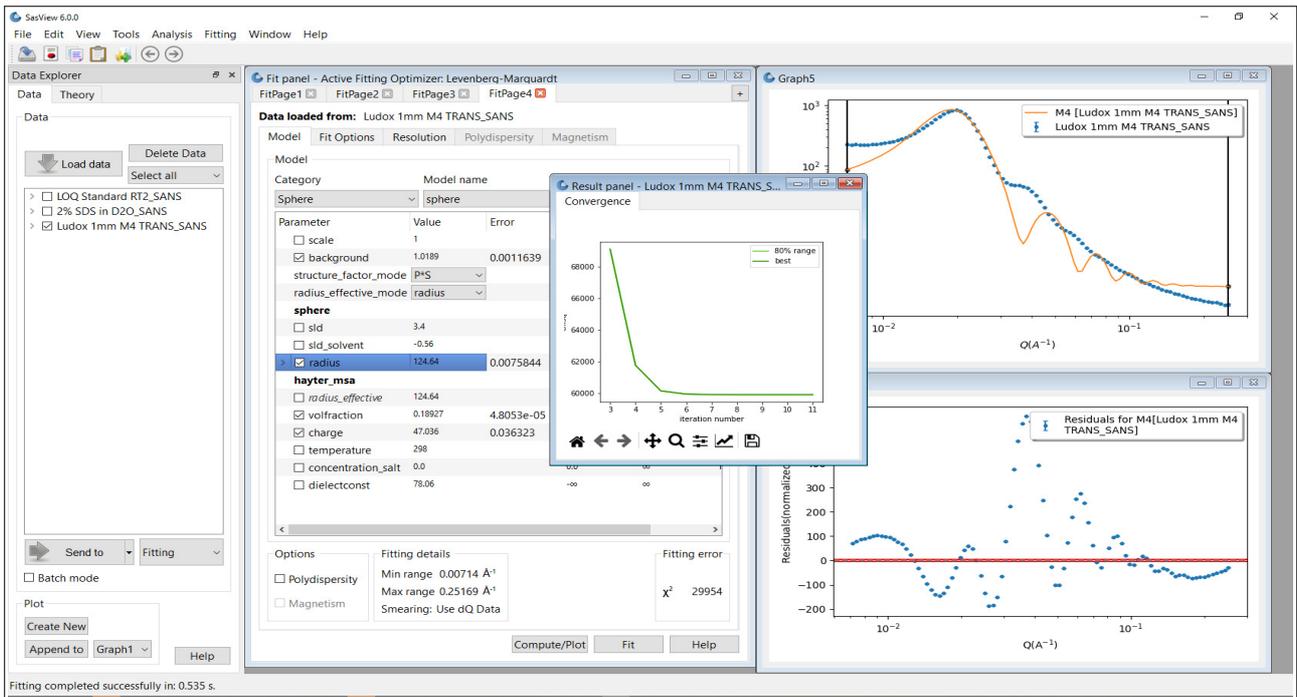
- *sld*: $3.4 \times 10^{-6} \text{ \AA}^{-2}$ (calculated)
- *sld_solvent*: $-0.56 \times 10^{-6} \text{ \AA}^{-2}$ (calculated)
- *volfraction*: 18 %_v (based on the manufacturers data sheet; 40 %_w)
- *temperature*: 298 K
- *concentration_salt*: 0 M
- *dielectconst*: 78.06 (Vidulich *et al*, *J Phys Chem*, 1967)

Enter these values and click **Compute/Plot**.



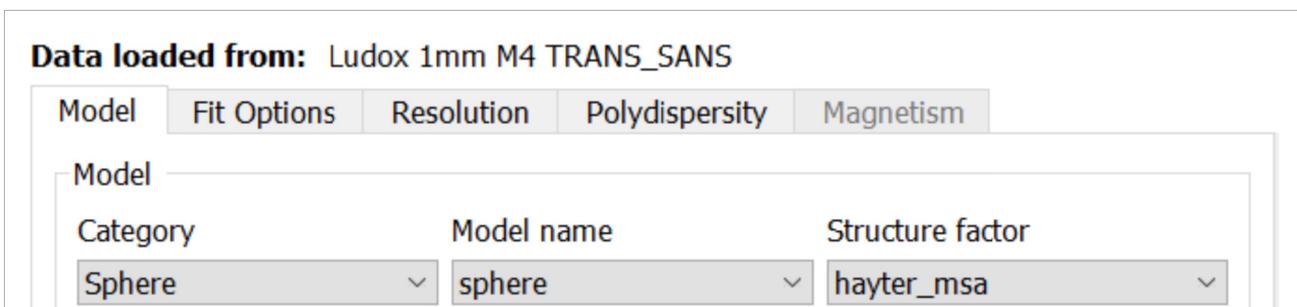
Clearly the particle *radius* is too small at present. Increase it to 150 Å.

Now check the *background*, *radius*, *volfraction*, and *charge* parameters and click **Fit**.

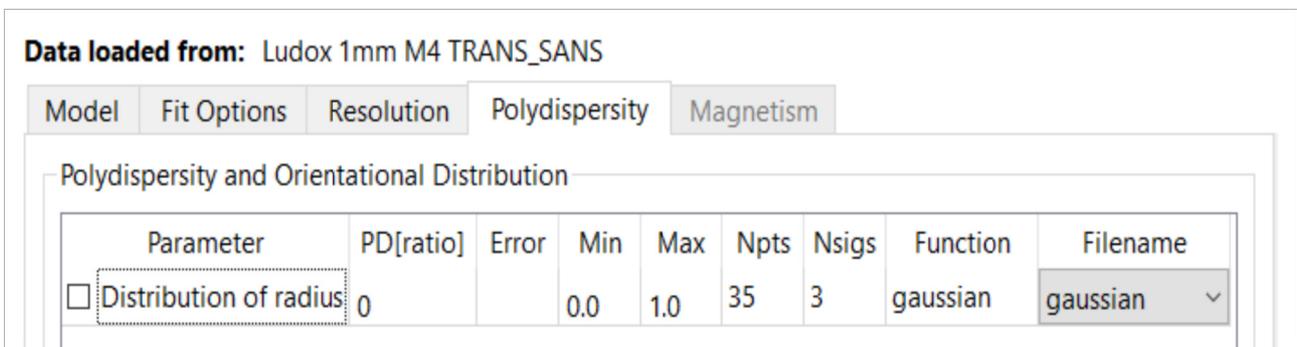


The problem now is that we are once again calculating a theory curve just for monodisperse particles. In Example 2 we managed to obtain a good solution without having to specifically address polydispersity, most likely because the size of the micelles meant any fringes in the scattering curve were beyond the range of the data.

In the lower part of FitPage, click on the checkbox named **Polydispersity**. This activates the tab named *Polydispersity*.



Click on this tab.



Particle size distributions are usually described by a Log-Normal distribution, so use the dropdown to change the *Function* to **lognormal**.

Tip: By default SasView always selects a Gaussian distribution. If you need a different distribution, select it before you set any distribution parameters. The SasView help documentation provides some guidance on appropriate use cases for different distributions.

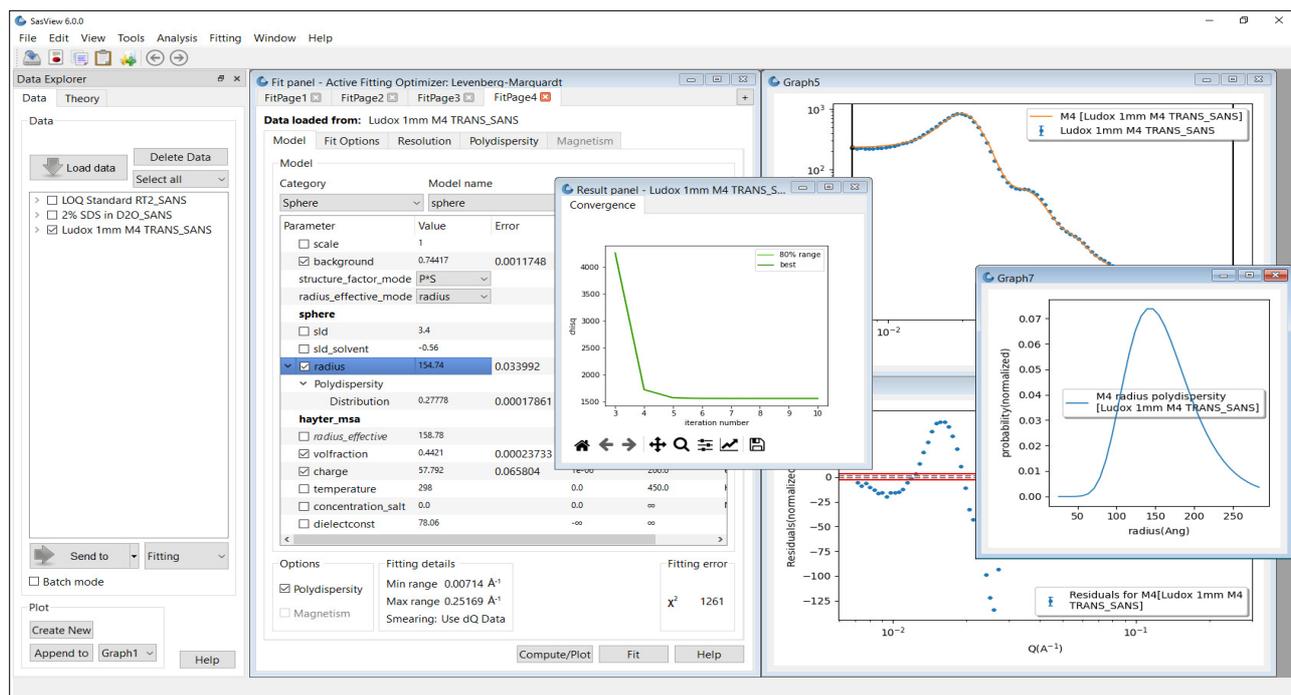
The width of the chosen polydispersity distribution is determined by the *PD* parameter and the definition of this changes depending on the distribution. For the Log-Normal distribution:

$$PD = \sigma_{normal}$$

where σ_{normal} is the width of the underlying Normal distribution. But whatever the distribution, $PD=0$ represents monodispersity (a very sharp distribution) and $PD=1$ represents an extremely broad distribution.

Enter $PD=0.15$ and click **Fit**.

Now check *Distribution of radius* and click **Fit** again.

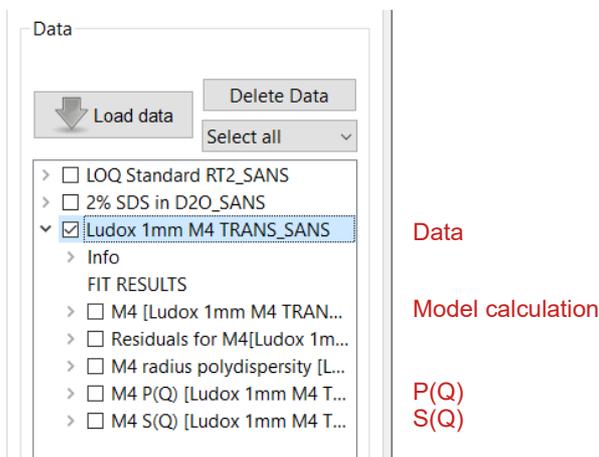


Whilst there is certainly room for improvement in both the *Reduced Chi2* and *Residuals*, this is nonetheless a promising fit to the data. The *radius* is within the size range typical of this material, but the *volfraction* is higher than anticipated. Seeming inconsistencies like this should always trigger a re-evaluation of the priors. For example, although Ludox® dispersions are described as being in water, is the dispersion medium truly water? Or does it contain residual components from the manufacturing process which may make the assumptions used about the values of *slid_solvent*, *concentration_salt* and *dielectconst* less valid?

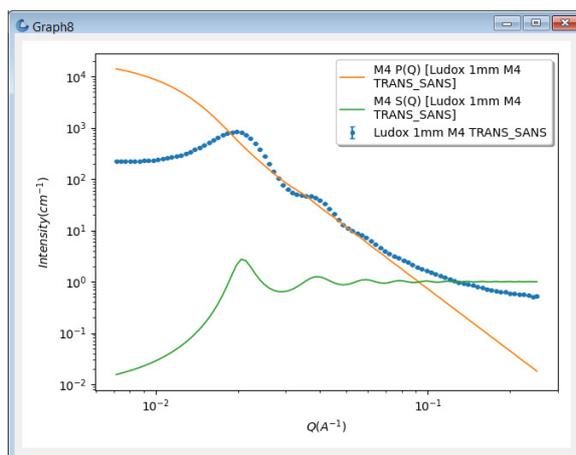
Also notice that an additional graph window has appeared depicting the actual form of the Log-Normal size distribution used in the model calculation.

Another nice feature is that it is also possible to see the separate contributions that the P(Q) and S(Q) make to the overall result.

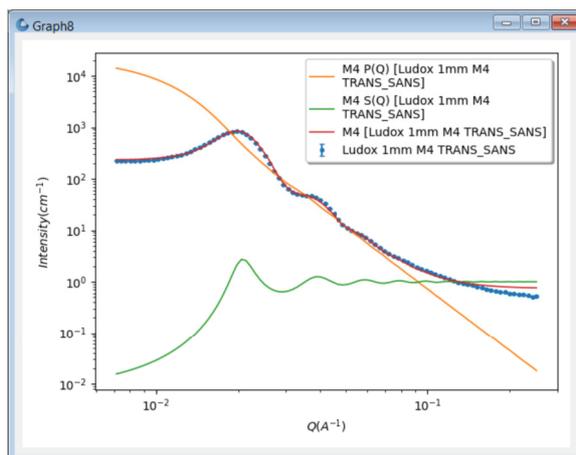
In the *Data Explorer*, click the > symbol to expand the Ludox® entry.



Check the P(Q) and S(Q) boxes and then click the **Create New Plot** button below.



If you also want to add the fit to this graph, at the top of the *Data Explorer* choose **Unselect all**, or just uncheck the boxes below already selected. Then select the model calculation dataset, and at the bottom of the SasView window select **Append to Graph8** and click the **Append to** button.



Further Information

For further information, please consult the

SasView Tutorial Series

or

<http://www.sasview.org>

or email

help@sasview.org